

Die Substanzen III–VIII wurden mit Hilfe von Hydrolysen, UV, NMR (vgl. Tab. 1) und z.T. Chromatographie mit authentischen Substanzen als Rutin (III), Isoquercitrin (IV), Isorhamnetin-3-*O*-rutosid (V), Isorhamnetin-3-*O*-glucosid (VI), Albosid [4] (VII) und Isorhamnetin (VIII) identifiziert. Die Hauptmenge machten V und VI aus.

Vom biogenetischen Standpunkt bilden die Substanzen III–VII eine interessante Reihe, bestehend aus den Glucosiden und Rhamnoglucosiden von Quercetin (IV, III) und dessen Mono- (VI, V) und Dimethyläther (VII); nur das Rutosid des Dimethyläthers konnte nicht nachgewiesen werden.

Durch Chromatographie der isolierten blau fluoreszierenden Verbindungen mit authentischen Substanzen und UV wurden Chlorogen-, Isochlorogen-, Ferula- und Gentisinsäure, sowie Äsculetin und Äsculin identifiziert. Aus Aceton-Extrakten wurden die folgenden Substanzen isoliert und durch Chromatographie identifiziert: Catechin, Epicatechin (Hauptsubstanzen), die Proanthocyanidine B2, B4, neben wenig B1 und B3 (Struktur, siehe Lit. [5]). Dadurch unterscheidet sich *S. alba* von *S. caprea* und *S. viminalis*, die als Hauptsubstanzen die Proanthocyanidine B3 und C2 enthalten [5].

EXPERIMENTELLES

Das Pflanzenmaterial, das in der Nähe von Gmünd geerntet wurde, wurde von Dr. R. Müller, Weleda, auf seine Identität überprüft. Belegexemplare befinden sich bei uns. Die NMR-Spektren wurden von den TMSi-Derivaten in CCl₄ auf einen Jeol JNM-C-60 HL Instrument (VI, VII) oder einen Bruker HFX 90 Gerät (I–III, V) mit TMS als innerem Standard aufgenommen. *R_f*-Werte wurden durch DC auf Cellulose mit CHCl₃–HOAc–H₂O (55:45:10) als Fließmittel bestimmt.

Isolierung der Flavonoide. Die frischen Blätter (22 kg) wurden mit MeOH kalt extrahiert. Die eingedampften Extrakte wurden nach Aufnehmen in heißem H₂O durch Extraktion mit versch. Lsg.-Mittel in 4 Frakt. aufgeteilt. Diese Frakt. wurden durch präparative PC (HOAc 15%) in 14 Zonen aufgeteilt. Diese Zonen wurden nach Elution mit MeOH chromatographiert an Sephadex LH-20 (70% MeOH), wodurch u.a. I und II als reine Stoffe gewonnen werden konnten.

Terniflorin (I), Schmp. 267° (MeOH) (Lit. [3] 266–7° (Zers.)). *R_f* 0,49. UV (MeOH), $\lambda_{\max}$ nm: 269, 319; (NaOMe): 272, 305,

367; (MeOH–AlCl₃): 276, 301, 321, 381; (AlCl₃–HCl): 278, 300, 322, 375; (NaOAc): 268, 325, 383; (NaOAc–H₃BO₃): 269, 321. IR (KBr), $\nu_{\max}$ cm⁻¹: 1690 (O=C–(C=C)–OR) (Lit. [3] 1686).

Apigenin-7-*O*-(4-*O*-*p*-cumarylglucosid) (II), Schmp. 239–42° (MeOH). *R_f* 0,63. UV (MeOH), $\lambda_{\max}$ nm: 268, 333; (NaOMe): 275, 327, 391; (MeOH–AlCl₃): 276, 303, 348, 384; (AlCl₃–HCl): 277, 301, 343, 382; (NaOAc): 276, 385; (NaOAc–H₃BO₃): 269, 344. IR (KBr), $\nu_{\max}$ cm⁻¹: 1690 (O=C–(C=C)–OR).

3',7-Dimethylquercetin-3-*O*-glucosid (Albosid) (VII), Schmp. 179–82° (MeOH) (Lit. [4] 178–80°). *R_f* 0,75.

Hydrolyse von I. 2 mg I wurden bei 100° mit 3 ml 1 N HCl (H₂O–MeOH (1:1)) 20 min. hydrolysiert.

Methanolyse von I. 5 mg I wurden bei Raumtemp. mit 4 ml 0,2% NaOMe–MeOH versetzt. Proben wurden stündlich entnommen und DC untersucht.

Umbildung von II in I. 2 mg II wurden in 2 ml MeOH gelöst und bei Raumtemp. 4 Wochen stehengelassen. Zunächst täglich, dann wöchentlich wurden Proben entnommen und DC untersucht (nach 4 Wochen 50%ige Umbildung).

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LITERATUR

1. Harborne, J. B. and Williams, C. A. (1975) in *The Flavonoids* (Harborne, J. B., Mabry, T. J. and Mabry, H., eds), pp. 376–441. Chapman & Hall, London.
2. Wagner, H. (1974) *Fortschr. Chem. Org. Naturst.* **31**, 153.
3. Aritomi, M. (1963) *Chem. Pharm. Bull.* **11**, 1225.
4. Thieme, H. (1968) *Tetrahedron Letters* 2301.
5. Thompson, R. S., Jacques, D., Haslam, E. and Tanner, R. J. N. (1972) *J. Chem. Soc. Perkin Tran.* **1**, 1387.
6. Birkofer, L., Kaiser, C., Hillges, B. and Becker, F. (1969) *Justus Liebigs Ann. Chem.* **725**, 196.
7. Birkofer, L., Kaiser, C. and Thomas, U. (1968) *Z. Naturforsch. B23*, 1051.
8. Birkofer, L., Kaiser, C., Kosmol, H., Romussi, G., Donike, M. and Michaelis, G. (1966) *Justus Liebigs Ann. Chem.* **699**, 223.

Phytochemistry, 1976, Vol. 15, pp. 1085–1086. Pergamon Press. Printed in England.

NEW FLAVONOIDS FROM *FLOURENSIA CERNUA*

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In connection with a biosystematic investigation of the genus *Flourensia*, five 6,8-di-*C*-glycosylflavones and two methoxylated flavonols have been isolated and identified from leaf material of *Flourensia cernua* DC. In addition,

two previously reported [1] methoxylated flavones, cirsimaritin and hispidulin were also characterized. *Flourensia cernua* (commonly called tarbrush, black brush or ojasen) is a frequent member of the desert vegetation

throughout much of the Chihuahuan Desert Region of north-central Mexico and southwestern United States.

The major component of the aqueous methanol-soluble material was a complex mixture of no less than five 6,8-di-*C*-glycosylapigenins: 6,8-di-*C*-glucosylapigenin (vicenin-2) (1), 6-*C*-xylosyl 8-*C*-glycosylapigenin (vicenin-1) (2), 6-*C*-arabinopyranosyl 8-*C*-glucosylapigenin (isoschaftoside) (3), 6-*C*-glucosyl 8-*C*-arabinopyranosylapigenin (schaftoside) (4) and 6-*C*-glucosyl 8-*C*-arabinosylapigenin (neoschaftoside) (5). 1, 3 and 4 were isolated in crystalline form and unambiguously identified by IR, MS and co-chromatography. To our knowledge, this is the first identification of isoschaftoside as a natural compound. Previously, it had been obtained by acid treatment of natural schaftoside [2]. Compound (2) was found as a by-product in the mother liquors of (3). Its presence was detected by TLC of the permethylation product of these mother liquors, PM (2) and PM (3) being well separated. The MS of PM (2) was identical with that of PM (vicenin-1). Then TLC of the mother liquors on activated silicagel in APEM P 20 allowed the confirmation of (2).

Compound (5) previously isolated from *Catananche caerulea* [3] and then misnamed isoschaftoside, is an isomer of schaftoside in which arabinose could be in the furanose form. Heating schaftoside with acids led to a mixture of (3), (4), and (5). (5) appeared as the faster migrating band on PC in BAW. Elution of this band gave a single spot (in BAW) with the same R_f as (5). TLC of the permethylation product of the eluate indicated the presence of three bands, all giving MS characteristic for PM 6-*C*-hexosyl 8-*C*-pentosylapigenins, one of these bands co-chromatographing with PM (5).

The CHCl_3 extraction yielded four flavonoid methyl ethers: 3-*O*-methylquercetin, 3, 7-di-*O*-methylkaempferol, 5,4'-dihydroxy-6,7-dimethoxyflavone (cirsimaritin) and 5,7,4'-trihydroxy-6-methoxyflavone (hispidulin). These were isolated and identified by UV, NMR and TLC comparison with authentic samples. Spectral values and color reactions for these compounds were identical with previously reported values [1,4,5].

EXPERIMENTAL

Leaf material was collected near Alpine, Texas (Brewster Co.) and a voucher specimen is deposited in the University of Texas at Austin, Texas (Bacon and Hartman 1443). All 2-D

chromatograms were on Whatman 3MM paper and were developed first in TBA (*t*-BuOH-HOAc- H_2O , 3:1:1) and then in 15% HOAc. NMR spectra were recorded using tetramethylsilane as an internal standard. Preparation of TMS ethers, permethylation products and TLC were carried out by standard procedures [5,6]. Air-dried and powdered leaves (150 g) of *Flourensia cernua* were extracted exhaustively with CHCl_3 . Combined extracts were taken to dryness *in vacuo*, yielding a dark green syrup (10.5 g). This syrup was chromatographed over polyamide (200 g packed in elution solvent); the column was initially washed with CHCl_3 -EtOAc (3:1) and later CHCl_3 -MeOH-MeEtKetone (12:3:1). Bands were collected in fractions, and purified by TLC in appropriate solvents. The four methylated flavonoids were identified from the CHCl_3 -soluble material. After drying to remove residual CHCl_3 , the leaf material was re-extracted with aq 85% MeOH. The extract was taken to dryness *in vacuo*, yielding a dark brown syrup (5 g). This was taken up in H_2O and partitioned with CHCl_3 . The aq portion was again taken to dryness and chromatographed over polyamide (200 g packed in 90% MeOH), eluting with progressively more aq MeOH. Bands were collected as fractions and purified by rechromatographing over polyamide (50 g packed in 100% MeOH). One band from the column yielded compounds 1-5 as a mixture of di-*C*-glycosylapigenins (from UV and R_f). MS of the permethylated mixture showed it to contain di-*C*-hexosyl and *C*-pentosyl *C*-hexosyl apigenins in the ratio (1:3). Cellulose column chromatography of the mixture in 15% HOAc directly gave crystalline (3) from the last fractions, (2) being present in the mother liquors. The first fractions contained a mixture of (1), (4) and (5) which were separated by repeated PC in BAW (4:1:5).

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REFERENCES

1. Rao, M. N., Kingston, D. G. I. and Spittler, T. D. (1970) *Phytochemistry* **9**, 227.
2. Chopin, J., Bouillant, M. L., Wagner, H. and Galle, K. (1974) *Phytochemistry* **13**, 2583.
3. Proliac, A., Raynaud, J., Combier, H., Bouillant, M. L. and Chopin, J. (1973) *C. R. Acad. Sci. Paris*, **277D**, 2813.
4. Valesi, A. G., Rodriguez, E., Vander Velde, G. and Mabry, T. J. (1972) *Phytochemistry* **11**, 2821.
5. Mabry, T. J., Markham, K. R. and Thomas, M. B. (1970) *The Systematic Identification of Flavonoids*, Springer-Verlag, Berlin.
6. Bouillant, M. L., Favre-Bonvin, J. and Chopin, J. (1975) *Phytochemistry* **14**, 2267.

A NEW FLAVONOID FROM *PLUCHEA SAGITTALIS**

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Plant. *Pluchea sagittalis* (Lam.) Cabrera (Compositae) is a perennial shrub that grows in Northeastern Argentin-

tina (local name "lucera" or "yerba del lucero") [1]. *Source*. Aerial parts were collected at Concepción del Uruguay, Province of Entre Rios, Argentina. A voucher specimen is deposited in the University Herbarium (Museo de Botánica, Universidad de Buenos Aires,

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